

INFRARED NEURAL STIMULATION MECHANISM MODELING

1 Supervising staff

Antoine Nonclercq (antoine.nonclercq@ulb.be), Louis Vande Perre (louis.vande.perre@ulb.be).

2 Background

Neurostimulation is widely used to treat chronic conditions. It is usually performed by electrical stimulation of tissues. However, it suffers from several shortcomings, including stimulation artifacts, low spatial resolution, incompatibility with magnetic resonance imaging, and the order in which nerve fibers are activated is not physiological. Optical stimulation could overcome all these shortcomings.

Among the optical nerve stimulation techniques, infrared neural stimulation (INS) is very promising because it does not require any prior genetic modification. INS consists of nerve activation due to transient exposure to infrared light. Our team has developed an experimental setup capable of compound action potential (CAP) elicitation in an excised rat sciatic nerve with the help of INS [1], [2]. The setup simultaneously records the nerve electrophysiological activity and the temperature at the surface of the nerve to link the CAP triggering and temperature gradients. However, numerical modeling would be helpful to investigate further why heat accumulation results in larger CAP amplitude. Moreover, modeling the light diffusion could help us better monitor the thermal gradients induced in the nerve based on the surface temperature measurements.

In the literature, three main hypotheses are currently considered as the underlying mechanism of INS: membrane capacitance change [3]–[7], membrane nanoporation [8], [9] or thermosensitive ion channels [10], [11] could be responsible for the axon depolarization. Therefore, the precise modeling of INS is complex, but numerous axon modeling could be used to explore the impact of temperature on the nerve cell. For the sake of simplicity, this project could rely on the Hodgkin-Huxley (HH) model, extensively used in neuroscience, which has already been adapted in the context of the INS [12]–[14]. However, Forrest argues that this simplistic model does not conveniently include thermodynamics considerations in action potential triggering [15]. Therefore, another model than the one developed by Hodgkin and Huxley might be considered.

Regarding the heat diffusion in the nerve during INS, previous work already investigated how temperature evolves at different spots in the nerve [16], [17]. However, these models do not fully describe the type of thermal gradients we observed in our experiments. Based on these works, a new model could be developed to analyze the results of our ex-vivo experiments better.

Experimental results

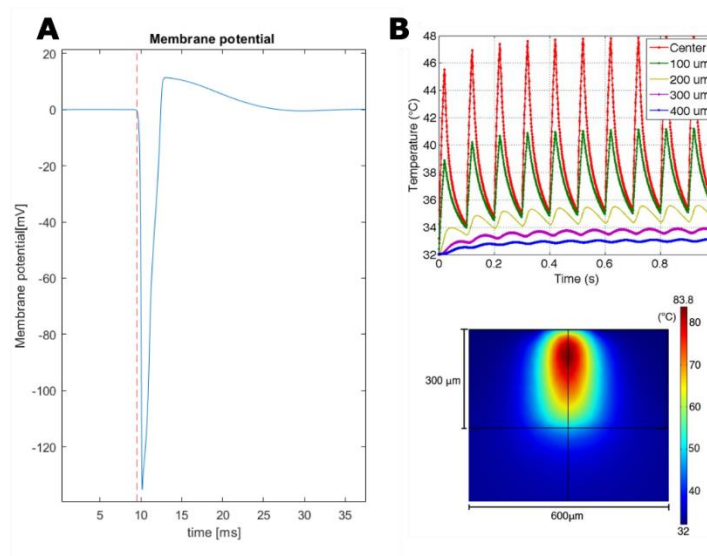
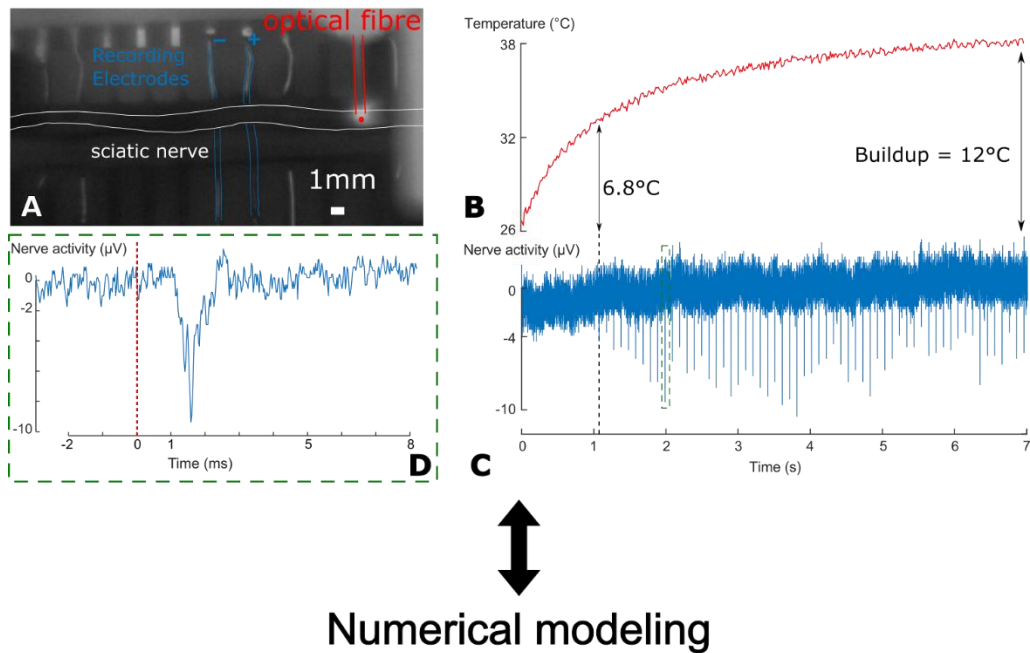


Fig. 1 Experimental results Infrared neural stimulation performed on an ex vivo sciatic nerve. (A) Grayscale image of the nerve obtained with IR camera. Brighter pixel is hotter. (B) Temperature of the center of the laser spot highlighted in (A) along the time. (C) Electrophysiological recording of the nerve. (D) Zoom on the 10th action potential evoked by the light pulse train after 2 seconds of stimulation (see green dashed box on C).

Numerical modeling (A) Action potential evoked with a basic Hodgkin-Huxley axon model. (B) Modeling of heating during INS (from [17]).

3 Work

The main goal of this project is to reuse the existing axon model or develop a similar model to investigate the impact of a rapid temperature increase on the membrane. The action potential (AP) elicitation will be based on a change in membrane capacitance, at least for the first model iteration. In parallel, modeling

of the heat diffusion - to better quantify the spatiotemporal temperature gradients in our experiments - could be achieved.

This work is exploratory and will require creativity and autonomy.

4 Bibliography

- [1] L. vande Perre *et al.*, “A Setup for Conduction Velocities and Temperature Gradients Measurements during Infrared Neurostimulation,” in *2022 IEEE Biomedical Circuits and Systems Conference (BioCAS)*, 2022, pp. 453–457. doi: 10.1109/BioCAS54905.2022.9948671.
- [2] J. Cury *et al.*, “Infrared neurostimulation in ex-vivo rat sciatic nerve using 1470 nm wavelength,” *J Neural Eng*, vol. 18, no. 5, p. 056018, 2021, doi: 10.1088/1741-2552/abf28f.
- [3] B. I. Pinto, C. A. Z. Bassetto, and F. Bezanilla, “Optocapacitance: physical basis and its application,” *Biophys Rev*, vol. 14, no. 2, pp. 569–577, 2022, doi: 10.1007/s12551-022-00943-9.
- [4] M. Plaksin, E. Shapira, E. Kimmel, and S. Shoham, “Thermal Transients Excite Neurons through Universal Intramembrane Mechanoelectrical Effects,” *Phys Rev X*, vol. 8, no. 1, p. 11043, 2018, doi: 10.1103/PhysRevX.8.011043.
- [5] M. G. Shapiro, K. Homma, S. Villarreal, C. P. Richter, and F. Bezanilla, “Infrared light excites cells by changing their electrical capacitance,” *Nat Commun*, vol. 3, no. March, pp. 310–376, 2012, doi: 10.1038/ncomms1742.
- [6] Z. Ebtehaj, A. Hatef, M. Malekmohammad, and M. Soltanolkotabi, “Computational Modeling and Validation of Thermally Induced Electrical Capacitance Changes for Lipid Bilayer Membranes Irradiated by Pulsed Lasers,” *Journal of Physical Chemistry B*, vol. 122, no. 29, pp. 7319–7331, 2018, doi: 10.1021/acs.jpcc.8b02616.
- [7] G. Bondelli *et al.*, “Shedding Light on Thermally Induced Optocapacitance at the Organic Biointerface,” *Journal of Physical Chemistry B*, vol. 125, no. 38, pp. 10748–10758, 2021, doi: 10.1021/acs.jpcc.1c06054.
- [8] H. T. Beier, G. P. Tolstykh, J. D. Musick, R. J. Thomas, and B. L. Ibey, “Plasma membrane nanoporation as a possible mechanism behind infrared excitation of cells,” *J Neural Eng*, vol. 11, no. 6, 2014, doi: 10.1088/1741-2560/11/6/066006.
- [9] C. C. Roth, R. A. Barnes, B. L. Ibey, R. D. Glickman, and H. T. Beier, “Short infrared (IR) laser pulses can induce nanoporation,” *Clinical and Translational Neurophotonics; Neural Imaging and Sensing; and Optogenetics and Optical Manipulation*, vol. 9690, p. 96901L, 2016, doi: 10.1117/12.2214892.
- [10] E. S. Albert *et al.*, “TRPV4 channels mediate the infrared laser-evoked response in sensory neurons,” *J Neurophysiol*, vol. 107, no. 12, pp. 3227–3234, 2012, doi: 10.1152/jn.00424.2011.
- [11] E. Suh, A. D. Izzo, M. Otting, J. T. Walsh, and C. Richter, “Optical stimulation in mice lacking the TRPV1 channel,” *Proceedings of SPIE - The International Society for Optical Engineering*, vol. 7180, no. February, pp. 71800S–1, 2009, doi: 10.1117/12.816891.
- [12] S. Fribance, J. Wang, J. R. Roppolo, W. C. de Groat, and C. Tai, “Axonal model for temperature stimulation,” *J Comput Neurosci*, vol. 41, no. 2, pp. 185–192, 2016, doi: 10.1007/s10827-016-0612-x.
- [13] M. J. Alemzadeh-Ansari, M. A. Ansari, M. Zakeri, and M. Haghjoo, “Influence of radiant exposure and repetition rate in infrared neural stimulation with near-infrared lasers,” *Lasers Med Sci*, vol. 34, no. 8, pp. 1555–1566, 2019, doi: 10.1007/s10103-019-02741-4.
- [14] C. A. Maldonado, B. D. Wooley, and J. J. Pancrazio, “The excitatory effect of temperature on the Hodgkin-Huxley model,” *Impulse: The Premier Undergraduate Neuroscience Journal*, pp. 1–7, 2015, [Online]. Available: [http://impulse.appstate.edu/sites/impulse.appstate.edu/files/Maldonado et al..pdf](http://impulse.appstate.edu/sites/impulse.appstate.edu/files/Maldonado%20et%20al..pdf)

- [15] M. D. Forrest, “Can the Thermodynamic Hodgkin-Huxley Model of Voltage-Dependent Conductance Extrapolate for Temperature?,” *Computation*, vol. 2, pp. 47–60, 2014, doi: 10.3390/computation2020047.
- [16] Zhou, R. et al. 2022. “Theoretical simulation of the selective stimulation of axons in different areas of a nerve bundle by multichannel near-infrared lasers.” *Medical and Biological Engineering and Computing*. 60, 1 (2022), 205–220. DOI:<https://doi.org/10.1007/s11517-021-02475-y>.
- [17] Liljemalm, R. et al. 2013. “Heating during infrared neural stimulation.” *Lasers in Surgery and Medicine*. 45, 7 (2013), 469–481. DOI:<https://doi.org/10.1002/lsm.22158>.